

# Characterization of soil fungal communities in lavender and oil-bearing rose plantations using DNA metabarcoding

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Soil fungi are key component of belowground biota characterized by great diversity and complex structure. They have different functional roles (e.g. decomposers, mutualists, pathogens, etc.) and can significantly affect plant and soil health. They are involved in all ecosystem processes and services but factors that influence their functioning are not well understood. The lack of knowledge is mainly related to their high diversity, hidden life mode and difficulty to culture. Therefore, culture-independent DNA-based identification methods for soil surveys are widely used. Recent studies use a high-throughput-sequencing (HTS) that allow an in-depth analysis of the entire fungal community composition in various environmental samples.



In the frame of the National Research Program "Healthy Foods for a Strong Bio-Economy and Quality of Life" we are studying the fungal diversity and composition in soils of lavender and oil-bearing roses managed by conventional and organic farming by applying DNA metabarcoding. We have two general objectives: 1) to examine the impact of cropping systems on soil fungal communities and 2) to propose management measures and agronomic practices for improving soil and plant health.



The study was conducted in 20 plantations located in the region of Kazanlak. Ten plots from each crop, managed by conventional and organic farming (2 x 5), were selected and sampled in the period of 2019-2020. Each soil sample is composed of 10 sub-samples collected from the plant rhizosphere at 0–5 cm depth. Soil was thoroughly mixed to homogenize and 5-10 g were stored for processing. Soil fungal composition was studied by sequencing the internal transcribed spacer 2 (ITS2) rDNA region. Fungal ITS2 amplicon library preparation and the sequencing were carried out using by the DNASense laboratory based on an Illumina protocol. Reads processing and classification were assigned to taxa using SINTAX algorithm and UNITE reference database v. 8.0 (Nilsson et al., 2018). The diversity and community structure were analysed at operational taxonomic units (OTUs) level using DNASense data analysis app.

Heatmap of the most abundant fungal OTUs and genera

Genus; OUT/Sample	LAVENDER										ROSE														
	13-LO	14-LO	15-LO	16-LC	17-LC	18-LC	19-LO2	19-LO3	21-LC	25-LO	26-LO	28-LC	07-RC	08-RC	09-RC	10-RO	11-RO	12-RO	20-RO	22-RC	23-RC	24-RO	27-RO/C		
g_Fusarium; Out1	3.4	11.2	4.0	4.6	5.7	6.3	9.0	9.6	6.9	5.9	5.2	10.1	4.7	1.0	5.2	7.4	3.8	3.7	0.3	0.7	4.9	10.9	6.5		
g_Cladosporium; Out2	1.7	5.4	2.6	3.4	2.3	2.4	1.7	3.4	4.2	4.3	1.8	3.5	3.1	9.1	1.8	1.6	1.3	2.0	10.7	1.1	10.8	1.3	1.6	3.6	
g_Fusarium; Out4	3.1	2.3	3.2	3.0	0.7	5.6	6.2	3.8	5.5	2.8	3.8	6.6	4.4	3.7	3.7	2.3	2.5	2.9	2.0	4.5	2.0	5.8			
g_Alternaria; Out3	1.0	1.5	0.9	0.9	1.6	1.1	1.1	0.5	1.1	1.1	2.8	2.7	0.9	3.7	12.4	3.4	0.5	0.5	1.3	0.3	26.9	1.8	2.1	2.6	
g_Phoma/Didymella; Out6	1.9	5.9	2.8	3.6	5.2	1.2	3.1	2.2	3.2	1.4	9.0	1.5	2.3	0.1	0.1	2.5	1.9	2.1	0.7	0.6	0.2	0.5	3.7	0.9	
g_Thelebolus; Out7	0.0	2.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
g_Albifimbria/Myrothecium; Out13	2.4	4.7	2.9	0.2	1.5	0.2	1.4	1.2	1.0	0.7	1.8	0.5	0.0	0.0	0.0	6.5	0.1	0.0	0.0	0.0	0.0	0.3	0.9	0.9	
g_Dothideomycetes; Out5	0.0	0.0	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
g_Robillarda; Out10	0.0	0.1	0.1	0.8	0.0	0.0	0.1	0.3	0.2	0.0	0.1	1.0	0.0	0.0	0.0	3.0	0.0	0.0	0.0	0.0	0.5	1.6	3.2	13.3	
g_Phoma/Didymella; Out21	0.3	1.4	3.8	1.7	2.4	0.8	1.8	0.6	0.6	1.1	0.8	0.2	0.1	0.2	0.5	1.9	0.3	0.7	0.8	0.2	0.3	1.9	0.6	0.8	
g_Truncatella; Out12	1.4	0.1	0.1	4.3	0.4	0.3	1.2	1.3	0.9	1.9	0.1	0.5	0.9	0.1	0.3	1.7	0.4	0.2	2.4	0.1	0.1	2.6	0.5	1.0	
o_Chaetothyriales; Out15	0.5	0.3	0.1	0.0	3.0	0.3	0.2	7.0	8.3	0.2	0.7	1.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
g_Naganishia; Out16	0.0	1.0	1.4	1.8	0.0	0.5	1.5	2.2	1.5	0.7	0.1	0.0	1.9	0.0	0.0	1.4	0.3	3.1	0.0	0.1	0.0	0.2	1.1	0.0	
g_Sollicozyma; Out14	0.0	1.6	2.3	0.5	0.0	0.7	1.0	2.0	1.4	1.8	0.2	0.0	1.8	0.0	0.0	1.4	0.1	0.6	0.0	0.0	0.5	0.3	1.2		
g_Mortierella; Out18	0.0	1.3	0.8	1.5	0.0	0.5	1.7	1.1	1.4	0.6	1.0	0.4	1.0	0.3	0.3	1.0	0.3	0.5	0.3	0.2	0.4	0.9	0.9	1.3	
g_Nothropoma; Out11	3.1	0.2	1.4	0.4	2.8	0.2	0.6	0.0	0.8	0.0	2.1	1.5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
g_Chaetothyriales; Out44	0.0	1.1	1.9	0.5	0.0	0.0	1.3	0.6	0.5	0.4	0.1	0.0	0.8	0.0	0.0	0.1	0.0	1.8	0.0	0.0	0.0	0.0	0.0	0.0	
g_Chaetosphaeronema; Out20	1.2	0.5	0.3	0.3	0.2	3.2	0.2	0.2	1.2	1.9	2.3	1.4	0.4	0.0	0.0	0.2	0.1	0.0	0.5	1.0	0.0	0.0	0.1	0.0	
g_Devriesia; Out24	0.1	1.9	2.3	0.6	0.0	0.2	0.6	1.3	0.5	0.3	0.0	1.1	0.1	0.1	1.1	0.5	1.1	0.2	1.0	0.0	0.1	0.4	0.0	0.0	
g_Minimedusa; Out31	3.1	0.2	1.7	0.3	1.6	0.0	0.3	0.0	0.7	0.4	1.3	0.3	1.0	0.2	0.3	0.4	0.3	0.0	0.0	0.1	0.1	1.7	0.7	0.0	
g_Cladosporium; Out19	0.3	0.3	0.2	0.5	0.5	0.8	0.2	0.2	0.7	0.5	0.2	0.9	0.3	1.0	0.8	3.3	0.3	0.1	2.1	0.1	0.7	0.2	0.1	0.5	
g_Stromatinia; Out37	0.7	1.2	0.2	4.4	0.0	0.1	1.2	1.1	1.4	0.1	0.2	0.0	0.3	0.0	0.0	2.1	0.0	0.2	0.0	0.0	0.0	0.6	0.6		
g_Sollicozyma; Out26	0.0	0.3	4.1	0.2	0.0	0.2	0.7	0.2	1.0	0.7	0.9	0.2	0.0	1.3	0.0	0.0	0.9	0.0	0.4	0.0	0.3	0.0	1.0	0.5	0.4
g_Rhizopus; Out29	0.6	0.2	0.5	0.3	0.1	0.1	1.3	1.0	1.9	0.9	1.3	1.3	0.4	0.0	0.0	1.3	0.2	0.0	0.0	0.0	0.2	0.4	0.8		
g_Phialemonium; Out17	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
g_Holtermanniella; Out30	0.0	0.5	0.8	0.3	0.0	1.6	0.9	1.3	1.8	0.1	0.0	0.2	0.6	0.8	0.3	0.6	0.7	0.5	0.0	0.0	0.0	0.0	0.6	0.1	
g_Saitozyme; Out25	0.0	0.9	0.8	0.0	0.0	0.0	0.9	1.2	0.8	2.5	0.0	0.0	0.8	0.0	0.0	0.1	0.0	0.4	0.0	2.2	0.0	0.1	0.0	0.3	
g_Aureobasidium; Out31	0.2	0.1	0.2	0.1	0.7	0.1	0.2	0.1	0.8	0.5	0.7	0.0	0.0	0.5	0.1	1.1	0.2	0							